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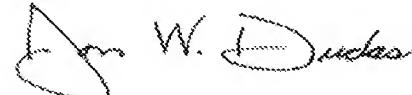
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APPLICATION NUMBER: 10/832,116

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042604
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U.S.PTO**UTILITY PATENT APPLICATION TRANSMITTAL
(Large Entity)**

(Only for new nonprovisional applications under 37 CFR 1.53(b))

Docket No.
34120USTotal Pages in this Submission
4**TO THE COMMISSIONER FOR PATENTS**

Mail Stop Patent Application
P.O. Box 1450
Alexandria, VA 22313-1450

Transmitted herewith for filing under 35 U.S.C. 111(a) and 37 C.F.R. 1.53(b) is a new utility patent application for an invention entitled:

**INHIBITION OF BIOGENIC SULFIDE PRODUCTION VIA BIOCIDE
AND METABOLIC INHIBITOR COMBINATION**

and invented by:

GARY E. JENNEMAN, ANNE GREENE, GERRIT VOORDOUW

If a **CONTINUATION APPLICATION**, check appropriate box and supply the requisite information:

Continuation Divisional Continuation-in-part (CIP) of prior application No.: _____

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Enclosed are:

Application Elements

1. Filing fee as calculated and transmitted as described below
2. Specification having 20 pages and including the following:

- a. Descriptive Title of the Invention
- b. Cross References to Related Applications (*if applicable*)
- c. Statement Regarding Federally-sponsored Research/Development (*if applicable*)
- d. Reference to Sequence Listing, a Table, or a Computer Program Listing Appendix
- e. Background of the Invention
- f. Brief Summary of the Invention
- g. Brief Description of the Drawings (*if filed*)
- h. Detailed Description
- i. Claim(s) as Classified Below
- j. Abstract of the Disclosure

15884 U.S.P.T.O. 10/8/2016
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UTILITY PATENT APPLICATION TRANSMITTAL
(Large Entity)

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Docket No.
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Application Elements (Continued)

3. Drawing(s) (*when necessary as prescribed by 35 USC 113*)
 - a. Formal Number of Sheets _____
 - b. Informal Number of Sheets _____
4. Oath or Declaration
 - a. Newly executed (*original or copy*) Unexecuted
 - b. Copy from a prior application (37 CFR 1.63(d)) (*for continuation/divisional application only*)
 - c. With Power of Attorney Without Power of Attorney
 - d. **DELETION OF INVENTOR(S)**
Signed statement attached deleting inventor(s) named in the prior application,
see 37 C.F.R. 1.63(d)(2) and 1.33(b).
5. Incorporation By Reference (*usable if Box 4b is checked*)
The entire disclosure of the prior application, from which a copy of the oath or declaration is supplied under Box 4b, is considered as being part of the disclosure of the accompanying application and is hereby incorporated by reference therein.
6. CD ROM or CD-R in duplicate, large table or Computer Program (Appendix)
7. Application Data Sheet (See 37 CFR 1.76)
8. Nucleotide and/or Amino Acid Sequence Submission (*if applicable, all must be included*)
 - a. Computer Readable Form (CRF)
 - b. Specification Sequence Listing on:
 - i. CD-ROM or CD-R (2 copies); or
 - ii. Paper
 - c. Statement(s) Verifying Identical Paper and Computer Readable Copy

Accompanying Application Parts

9. Assignment Papers (*cover sheet & document(s)*)
10. 37 CFR 3.73(B) Statement (*when there is an assignee*)
11. English Translation Document (*if applicable*)
12. Information Disclosure Statement/PTO-1449 Copies of IDS Citations
13. Preliminary Amendment
14. Return Receipt Postcard (MPEP 503) (*Should be specifically itemized*)
15. Certified Copy of Priority Document(s) (*if foreign priority is claimed*)
16. Certificate of Mailing
 First Class Express Mail (*Specify Label No.:* EL 993327444 US)

**UTILITY PATENT APPLICATION TRANSMITTAL
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Docket No.
34120US

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4

Accompanying Application Parts (Continued)

17. Additional Enclosures (*please identify below*):

Request That Application Not Be Published Pursuant To 35 U.S.C. 122(b)(2)

18. Pursuant to 35 U.S.C. 122(b)(2), Applicant hereby requests that this patent application not be published pursuant to 35 U.S.C. 122(b)(1). Applicant hereby certifies that the invention disclosed in this application has not and will not be the subject of an application filed in another country, or under a multilateral international agreement, that requires publication of applications 18 months after filing of the application.

Warning

An applicant who makes a request not to publish, but who subsequently files in a foreign country or under a multilateral international agreement specified in 35 U.S.C. 122(b)(2)(B)(i), must notify the Director of such filing not later than 45 days after the date of the filing of such foreign or international application. A failure of the applicant to provide such notice within the prescribed period shall result in the application being regarded as abandoned, unless it is shown to the satisfaction of the Director that the delay in submitting the notice was unintentional.

UTILITY PATENT APPLICATION TRANSMITTAL
(Large Entity)

(Only for new nonprovisional applications under 37 CFR 1.53(b))

Docket No.
 34120US

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18334 U.S. PTO
 042604

Fee Calculation and Transmittal

CLAIMS AS FILED

For	#Filed	#Allowed	#Extra	Rate	Fee
Total Claims	64	- 20 =	44	x \$18.00	\$792.00
Indep. Claims	4	- 3 =	1	x \$86.00	\$86.00
Multiple Dependent Claims (check if applicable)	<input type="checkbox"/>				\$0.00
				BASIC FEE	\$770.00
OTHER FEE (specify purpose)					\$0.00
				TOTAL FILING FEE	\$1,648.00

- A check in the amount of **\$1,648.00** to cover the filing fee is enclosed.
- The Director is hereby authorized to charge and credit Deposit Account No. **19-0522** as described below.
- Charge the amount of _____ as filing fee.
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 - Charge any additional filing fees required under 37 C.F.R. 1.16 and 1.17.
 - Charge the issue fee set in 37 C.F.R. 1.18 at the mailing of the Notice of Allowance pursuant to 37 C.F.R. 1.311(b).

Dated: April 26, 2004

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IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Application of :) Examiner:
JENNEMAN, GARY E. et al.) TC/A.U.:
Serial No.:) Confirmation No.:
Filed:) Customer No.: 23589
Title: INHIBITION OF SULFIDOGENIC)
BACTERIA VIA BIOCIDE)
AND METABOLIC INHIBITOR)
COMBINATION)

TRANSMITTAL

Transmitted herewith: Transmittal (4 pgs); Patent Application Specification (20 pgs); \$1,648 filing fee; Declarations/Powers of Attorney (3 pgs); IDS Transmittal (1 pg); IDS Statement w/accompanying references (34 pgs); Assignment Cover Sheet (1 pg); Assignment (1 pgs; \$40 recording fee; and return postcard. These documents are being deposited with the United States Postal Service "Express Mail Post Office to Addressee" service under 37 C.F.R. 1.10 in an envelope addressed to: Mail Stop Patent Application, Commissioner for Patents, P.O. Box 1450, Alexandria, VA, 22313-1450 on April 26, 2004.

EL 993327444 US
Express Mail No.

Respectfully submitted,

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**INHIBITION OF BIOGENIC SULFIDE PRODUCTION
VIA BIOCIDE AND METABOLIC INHIBITOR COMBINATION**

BACKGROUND OF THE INVENTION

5 **1. Field of the Invention**

The present invention relates generally to the control of biogenic sulfide production. In another aspect, the invention concerns the use of at least one biocide and at least one metabolic inhibitor to synergistically inhibit sulfide production by sulfate-reducing bacteria.

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2. Description of the Prior Art

The presence of sulfides (e.g., H₂S, HS⁻, and S²⁻) in fluids poses serious problems due to their toxicity, odor, and corrosive nature. It is well known that the presence of sulfides in many fluids is a consequence of the reduction of sulfates to sulfides by sulfate-reducing bacteria (SRB). SRB are routinely found in water associated with oil production systems and can be found in virtually all industrial aqueous processes including, for example, cooling-water systems, pulp and paper-making systems, chemical manufacturing, and petroleum refining.

20

Requirements for SRB activity and growth include a substantially anaerobic aqueous environment containing adequate nutrients, an electron donor, and an electron acceptor. A typical electron acceptor is sulfate, which produces H₂S upon reduction. A typical electron donor is a volatile fatty acid (e.g., acetic or propionic acids), although hydrogen can also function as an electron donor. Conditions in an oil reservoir subjected to seawater flooding are excellent for establishing SRB activity. Seawater contains a significant concentration of sulfate, while connate, or indigenous formation, water contains volatile fatty acids and other required trace nutrients (e.g., nitrogen and phosphorus). Conditions within industrial water systems, such as effluent streams from production operations or cooling water streams, are also conducive to SRB activity due to the anaerobic biofilm which is formed on pipeline, tank, or vessel walls. The same is true within the sewers and other piping and facilities associated with municipal wastewater handling systems.

Hydrogen sulfide (H₂S) is corrosive and reacts with metal surfaces to form insoluble iron sulfide corrosion products. In oilfield operations, H₂S partitions into the water, oil, and natural gas phases of produced fluids and creates a number of problems. For instance,

oil and gas that contain high levels of H₂S have a lower commercial value than low-sulfide oil and gas. Removing biogenic H₂S from sour oil and gas increases the cost of these products. In addition, H₂S is an extremely toxic gas and can be lethal to humans at even small concentrations. Its presence in wastewater systems poses a threat to worker safety. The
5 discharge of produced waters containing high levels of H₂S into aquatic or marine environments is hazardous because H₂S reacts with oxygen and lowers the dissolved-oxygen levels in the water.

Corrosion caused by SRB-produced H₂S frequently results in extensive damage. Pipe systems, tank bottoms, and other pieces of equipment can rapidly fail if they
10 have areas where microbial corrosion occurs. If a failure occurs in a pipeline or storage tank bottom, the released fluid can have serious environmental consequences. If a failure occurs in a high pressure water or gas line, the consequences may be worker injury or death. Any such failure involves substantial repair or replacement costs.

In the past there have been two main approaches to reducing the level of
15 sulfides in industrial fluids. One approach involved removing sulfides from the fluids after their formation. This post-formation removal approach, however, was frequently uneconomical or impractical, especially in oilfield operations. The other approach has been to treat the SRB-containing fluids with biocides or metabolic inhibitors to thereby kill or inhibit the growth of the SRB prior to significant biogenic sulfide formation. However, in
20 many instances high concentrations of biocides or metabolic inhibitors are required to effectively inhibit sulfide production by SRB. The costs associated with employing biocides or metabolic inhibitors in such high concentrations can be prohibitive.

OBJECTS AND SUMMARY OF THE INVENTION

It is, therefore, an object of the present invention to provide a method and
25 composition for more effectively and economically inhibiting biogenic sulfide production.

Another object of the present invention is to provide a composition that is effective to inhibit sulfide production by SRB at relatively low concentrations of the inventive composition.

It should be understood that the above-listed objects are only exemplary.
30 Further objects and advantages of the present invention will be apparent from the detailed description of the preferred embodiment, the claims, and the drawing figures.

Accordingly, one aspect of the present invention concerns a method of inhibiting sulfide production by SRB. The method comprises the steps of: (a) contacting the SRB with a first concentration of a biocide component, wherein the first concentration is less than about 90% of the minimum inhibitory concentration (MIC) of the biocide component; 5 and (b) contacting the SRB with a second concentration of a metabolic inhibitor component, wherein the second concentration is less than about 90% of the MIC of the metabolic inhibitor component.

Another aspect of the present invention concerns a method comprising 10 contacting SRB with a treated medium comprising an aldehyde and a metabolic inhibitor. The metabolic inhibitor is selected from the group consisting of nitrite, molybdate, and combinations thereof. The aldehyde and the metabolic inhibitor are present in the treated medium in an aldehyde to metabolic inhibitor molar ratio in the range of from about 50:1 to 15 about 1:50.

Still another aspect of the present invention concerns a composition for 20 effectively inhibiting sulfide production by SRB. The composition comprises: (a) a biocide component capable of directly killing a first portion of the SRB; and (b) a metabolic inhibitor component capable of inhibiting the sulfate-reducing growth of a second portion of the SRB without directly killing the second portion of the SRB. The biocide component is present in the composition in a first concentration that is less than about 90% of the MIC of the biocide 25 component. The metabolic inhibitor component is present in the composition in a first concentration that is less than about 90% of the MIC of the biocide component.

A further aspect of the present invention concerns a composition comprising 20 an aldehyde and a metabolic inhibitor selected from the group consisting of nitrite, molybdate, and combinations thereof. The aldehyde and the metabolic inhibitor are present in the composition in an aldehyde to metabolic inhibitor molar ratio in the range of from 25 about 50:1 to about 1:50.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENT

We have discovered that sulfide production by sulfate-reducing bacteria 30 (SRB) can be more effectively and economically inhibited by treating the SRB with certain synergistic combinations of biogenic sulfide inhibitors (BSIs). As used herein, "sulfate-reducing bacteria" or "SRB" shall denote one or more types of bacterium capable of facilitating the reduction of sulfates to sulfides. As used herein, "biogenic sulfide inhibitor"

or "BSI" shall be used as a generic term to denote any compound that effectively inhibits sulfide production by at least one type of sulfate-reducing bacterium. BSIs of particular significance in the present invention include biocides and metabolic inhibitors. As used herein, "biocide" shall denote a compound that directly kills at least one type of sulfate-reducing bacterium via contact therewith. As used herein, "metabolic inhibitor" shall denote a compound that effectively inhibits the sulfate-reducing activity of at least one type of sulfate-reducing bacterium, without directly killing the inhibited sulfate-reducing bacterium upon contact therewith. Metabolic inhibitors deprive SRB of the ability to produce ATP and, as a result, cells are unable to grow and/or divide. This inability to grow or divide 5 may eventually cause the death of some of the SRB; however, the cell death is not a direct 10 result of exposure to the metabolic inhibitors as it would be for biocides.

In accordance with one embodiment of the present invention, SRB are contacted with a treated medium comprising more than one BSI to thereby synergistically inhibit biogenic sulfide production. Preferably, the treated medium comprises at least one 15 biocide and at least one metabolic inhibitor. Biocides suitable for use in the present invention include both oxidizing and non-oxidizing biocides. Preferably, non-oxidizing biocides are employed. Suitable non-oxidizing biocides include, for example, aldehydes (e.g., formaldehyde, glutaraldehyde, and acrolein), amine-type compounds (e.g., quaternary amine compounds and cocodiamine), halogenated compounds (e.g., bronopol and 20 2-2-dibromo-3-nitrilopropionamide (DBNPA)), sulfur compounds (e.g., isothiazolone, carbamates, and metronidazole), and quaternary phosphonium salts (e.g., tetrakis (hydroxymethyl) phosphonium sulfate (THPS)). Metabolic inhibitors suitable for use in the present invention include, for example, nitrite, molybdate, tungstate, selenate, and anthraquinone. Other equivalent metabolic inhibitors for SRB may exist, but are not known 25 or foreseeable at the time of filing of this patent.

The synergistic inhibitory effect resulting from the combined use of more than one BSI (e.g., a biocide and a metabolic inhibitor) can be demonstrated by comparing the inhibitory effect of the combined BSIs with the inhibitory effect of each individual BSI, when used alone. This synergistic inhibitory effect can be quantified by comparing the 30 concentrations of the combined BSIs necessary to provide effective biogenic sulfide inhibition with the concentrations of each individual BSI necessary to provide effective sulfide inhibition when each individual BSI is used alone.

The concentration of an individual BSI necessary to effectively inhibit sulfide production by SRB can be expressed as a minimum inhibitory concentration (MIC). As used herein, "minimum inhibitory concentration" or "MIC" shall denote the minimum concentration of an individual BSI necessary to prevent sulfide production by SRB for 30 days after contact with the SRB is initiated. Each BSI has a unique MIC. For example, we have found that under certain test conditions, a 5 mM (milliMolar) concentration of glutaraldehyde (biocide) in a certain treated medium is the minimum concentration of glutaraldehyde necessary to prevent sulfide production by certain SRB for 30 days after the treated medium is first contacted with the SRB. Thus, under the conditions of this test, the 10 MIC of glutaraldehyde is 5 mM.

This patent uses the MIC of various BSIs as a reference to demonstrate that synergistic biogenic sulfide inhibition can be achieved when certain combinations of BSIs are employed at concentrations that are substantially less than the MIC of each individual BSI. Thus, the amount or concentration of a particular BSI used to treat the SRB can be 15 expressed as a percentage of the MIC of that particular BSI. It should be noted, however, that the MIC of a particular BSI can vary, depending upon numerous factors such as, for example, the type of SRB treated, the composition of the treated medium, and the temperature at which the SRB and treated medium are maintained. Thus, when SRB are treated with an amount of a particular BSI that is expressed as a percentage of the MIC for that BSI, it is assumed 20 that the MIC for that BSI was determined at the same conditions under which the SRB are currently being treated. For example, if a certain treated medium comprising glutaraldehyde and nitrite is used to treat certain SRB under certain conditions and the treated medium contains glutaraldehyde at 50% (by mole) of the MIC of glutaraldehyde, then the concentration of glutaraldehyde in the treated medium is one-half the concentration of 25 glutaraldehyde alone (i.e., without nitrite) in the treated medium that would be necessary to prevent sulfide production by the SRB for 30 days under the same conditions.

One embodiment of the present invention can be carried out by contacting SRB with at least one biocide and at least one metabolic inhibitor in either a simultaneous or sequential fashion. Preferably, the biocide and metabolic inhibitor components are 30 simultaneously contacted with the SRB by first combining the biocide (and/or a precursor of the biocide) and metabolic inhibitor (and/or a precursor of the metabolic inhibitor) in a treated medium and contacting the SRB with the treated medium. Nitrate is one example of a precursor of nitrite. The specific composition of the treated medium can vary greatly,

depending upon the particular application for which biogenic sulfide inhibition is sought. Thus, the treated medium can be any medium suitable for carrying the biocide and metabolic inhibitor components. Preferably, the treated medium is an aqueous-based medium, more preferably the treated medium comprises at least about 2% water by weight, more preferably at least about 50% water by weight, and most preferably at least 90% water by weight. The SRB with which the treated medium is contacted can reside in the treated medium itself or on a surface (e.g., the surface of a subterranean formation or the inner surface of a pipe or vessel) with which the treated medium comes into contact. In one application, the treated medium is brine (e.g., oilfield brine) that contains sulfates, SRB, a biocide, and a metabolic inhibitor. In certain instances, the biocide may be present as part of conventional oilfield chemicals, such as corrosion inhibitors. Thus, it may be preferred to employ biocides that exhibit other advantageous properties such as corrosion inhibition. For example, quaternary amines are good biocides and corrosion inhibitors.

The synergistic inhibition provided by the combined biocide and metabolic inhibitor components of the treated medium allow for effective biogenic sulfide inhibition at concentrations substantially less than the minimum inhibitory concentrations (MICs) of the individual components. Thus, it is preferred for the concentrations of the biocide and the metabolic inhibitor components of the treated medium to be less than the MICs of the individual biocide and metabolic inhibitor components. More preferably, the concentrations of both the biocide and the metabolic inhibitor are less than about 90% of their respective MICs. Still more preferably, the concentrations of one or both the biocide and the metabolic inhibitor are less than about 75% of their respective MICs. Even more preferably, the concentrations of one or both of the biocide and the metabolic inhibitor are less than about 50% of their respective MICs. Yet still more preferably, the concentrations of one or both the biocide and the metabolic inhibitor are less than about 35% of their respective MICs. Most preferably, the concentrations of one or both the biocide and the metabolic inhibitor are less than 25% of their respective MICs.

In a preferred embodiment of the present invention, the biocide is an aldehyde and the metabolic inhibitor is nitrite and/or molybdate. When the biocide is an aldehyde and the metabolic inhibitor is nitrite and/or molybdate it is preferred for the treated medium to have a biocide to metabolic inhibitor molar ratio in the range of from about 50:1 to about 1:50, more preferably about 20:1 to about 1:20, still more preferably about 10:1 to about 1:10, and most preferably 5:1 to 1:5. In addition, when the biocide is an aldehyde, it is

preferred for the concentration of the biocide in the treated medium to be in the range of from about 0.1 to about 5 mM, (milliMolar) more preferably about 0.1 to about 3 mM, and most preferably 0.1 to 2 mM. When the metabolic inhibitor is nitrite and/or molybdate, it is preferred for the concentration of the metabolic inhibitor in the treated medium to be in the range of from about 0.1 to about 5 mM, more preferably about 0.1 to about 3 mM, and most preferably 0.1 to 2 mM. In a particularly preferred embodiment of the present invention, the biocide component contacted with the SRB consists essentially of glutaraldehyde and the metabolic inhibitor component contacted with the SRB consists essentially of nitrite.

The treated medium and the SRB can be contacted in either an intermittent (i.e., batch) or continuous fashion. Preferably, the present invention is carried out in a substantially continuous manner. In either case, the concentrations of the biocide and metabolic inhibitor components, described above, are expressed as time-averaged concentrations. For example, if SRB is contacted with a treated medium in a batch mode having a frequency of once every 24 hours (1440 minutes), a duration of 14.4 minutes, and a batch concentration of 100 mM, the average concentration would be 1 mM (i.e., $100 \text{ mM} \times 14.4 \text{ min} / 1440 \text{ min}$). The following example is intended to be illustrative of the present invention and to teach one of ordinary skill in the art to make and use the invention. This example is not intended to limit the invention in any way.

EXAMPLE

In this Example, the effect of various biocide and metabolic inhibitor combinations and concentrations were investigated to determine their combined effect on sulfide production by SRB.

The sulfate-reducing bacterial (SRB) consortium used in this study was enriched from produced water obtained from the Coleville oil field near Kindersley, Saskatchewan, Canada. Serial enrichment in saline Postgate C medium (sPGC) resulted in a stable SRB consortium that was maintained for over one year prior to commencement of the biocide and metabolic inhibitor exposure experiments, described below. The SRB consortium was maintained by weekly transfer in sPGC medium, and incubated at 30°C. Saline Postgate C medium (sPGC) is a modification of medium C described in Postgate, J.R. The Sulfate-Reducing Bacteria. Cambridge: Cambridge University Press, pp. 30-34 (1984). The sPGC contained the following components per 1 liter of distilled water: 7 g NaCl; 1.2 g MgCl₂ 6H₂O; 0.5 g KH₂PO₄; 1 g NH₄Cl; 4.5 g Na₂SO₄; 0.042 g CaCl₂ 2H₂O; 0.03 g MgSO₄.

$7\text{H}_2\text{O}$; 0.004 g $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$; 0.28 g sodium citrate; 10 g 60% sodium lactate; 1 g yeast extract; and a trace amount of rezazurin.

The cultures used in this study were grown in 100 mL of modified Coleville synthetic brine medium (mCSB) in 160-ml serum bottles, with a headspace of 5% H_2 , 10% CO₂ balance N₂. The mCSB is described in Nemati M., Jenneman G.E., Voordouw G. *A mechanistic study on microbial control of souring in oil reservoirs*. Biotechnol. Bioeng. 74:424-434 (2001). The mCSB contained the following components per 950 milliliters of distilled water: 7 g NaCl; 0.027 g KH_2PO_4 ; 0.02 g NH_4Cl ; 0.24 g $\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$; 0.68 g $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$; 1 g $(\text{NH}_4)_2\text{SO}_4$; 0.68 g sodium acetate; 5.6 g sodium lactate syrup (60% v/v); 1.9 g NaHCO_3 ; and 50 ml micronutrient solution. The micronutrient solution contained the following components per 990 ml of distilled water: 2 g nitrilotriacetic acid; 0.006 g FeCl_3 ; 1.2 g $\text{CaSO}_4 \cdot 2\text{H}_2\text{O}$; 2 g $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$; 0.16 g NaCl; 1.4 g Na_2HPO_4 ; 0.72 KH_2PO_4 ; and 10 ml trace element solution. The 10 ml trace element solution contained the following components: 0.5 ml H_2SO_4 ; 2.28 g $\text{MnSO}_4 \cdot \text{H}_2\text{O}$; 0.5 g $\text{ZnSO}_4 \cdot 7\text{H}_2\text{O}$; 0.5 g H_3BO_3 ; 0.025 g $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$; 0.025 g $\text{NaMoO}_4 \cdot 2\text{H}_2\text{O}$; and 0.045 g $\text{CoCl}_2 \cdot 6\text{H}_2\text{O}$.

A 2% inoculum of freshly grown Coleville SRB enrichment was used in all cases. After inoculation, cultures were incubated overnight at 30°C, until the produced sulfide in the cultures was approximately 5 milliMolar (mM) (maximum concentration of produced sulfide in these cultures is approximately 12 mM). At this time, the biocide/metabolic inhibitor combinations were added. Cultures were incubated for 1 month after the addition of the biocide and metabolic inhibitor. If sulfate reduction and sulfide production resumed during the 1-month incubation period, inhibition was deemed unsuccessful.

Sulfate was measured using the turbidimetric method described in: American Public Health Association, Standard Methods for the Examination of Water and Wastewater. Washington, DC: American Water Works Association and Water Pollution Control Federation, pp. 439-440 (1992) as modified by the method described in Nemati M., Jenneman G.E., Voordouw G., *A mechanistic study on microbial control of souring in oil reservoirs*, Biotechnol. Bioeng. 74:424-434 (2001). Sulfide was analyzed by the colorimetric method described in Cord-Ruwisch, R., *A quick method for determination of dissolved and precipitated sulfides in cultures of sulfate-reducing bacteria*, J. Microbiol. Meth. 4:33-36 (1985). Nitrite was evaluated by the colorimetric method described in Nemati M., Jenneman G.E., Voordouw G., *A mechanistic study on microbial control of souring in oil reservoirs*, Biotechnol. Bioeng. 74:424-434 (2001). Cell growth was not monitored; the optical density

and color of various cultures changed significantly upon addition of some biocides or inhibitors, which interfered with optical density readings.

Various combinations of biocides and metabolic inhibitors were tested. For the purpose of this example, biocides are defined as agents that kill microorganisms directly.

- 5 The two metabolic inhibitors tested are both specific to SRB and are known to interfere with different stages of sulfate reduction to sulfide. Inhibition of sulfate reduction deprives SRB of the ability to produce ATP (the cellular energy currency), thus cells are unable to grow or divide and may eventually die, however cell death is not necessarily a result of exposure to these compounds, particularly at low concentrations where energy production might be
10 decreased but not completely inhibited.

For each biocide and metabolic inhibitor tested, the minimum inhibitory concentration (MIC; the minimum amount of biocide required to inhibit sulfate reduction and sulfide production in the SRB culture for one month) was determined. Combinations of pairs of biocides were tested at various concentrations to determine the MICs of several concentrations of each when mixed. The effectiveness of various biocide combinations was evaluated. Biocide combination effects were separated into five categories: antagonistic (one biocide had a negative effect on another such that more than the MIC for one biocide alone plus the second biocide at any amount was required for inhibition), additive (e.g., inhibition requires 25% of the MIC of one biocide and 75% of the other, or vice versa), indifferent, less than additive (more than an additive amount of the pair of biocides, but less than the MIC of each, is required for inhibition) or synergistic (less than an additive amount of the pair of biocides is required for inhibition).

The metabolic inhibitors evaluated were molybdate and nitrite. Six nonoxidizing biocides were evaluated alone and in combination with nitrite or molybdate (oxidizing biocides were not considered in this study). Both glutaraldehyde and formaldehyde are aldehyde-type biocides. Benzalkonium chloride is a representative of the quaternary amine group of biocides. Combinations of quaternary amine biocides and glutaraldehyde are commercially available for use in oilfield and other situations. Cocodiamines are from the amine and diamine biocide group. The cocodiamine biocide used in this study was T-397, provided by Brenntag Canada. Bronopol (2-bromo-2-nitropropane-1,3-diol) is a halogenated biocide. Tetrakis(hydroxymethyl)phosphonium sulfate (THPS) is a quaternary phosphonium salt. Biocides from several groups commonly used in oil field situations were purposely

chosen in order to allow a general evaluation of the effectiveness of each group when combined with specific metabolic inhibitors.

The test results for combinations of various biocides with the metabolic inhibitors (nitrite or molybdate) are shown in FIGS 1-10. In FIGS. 1-10, the open triangles (Δ) represent concentrations that did not successfully inhibit sulfide production for a full month, while the solid diamonds (◆) represent concentrations that successfully inhibited sulfide production for a full month. The diagonal line in each plot represents what the inhibitory concentrations would be if the biocide and metabolic inhibitor had a purely additive effect. Thus, successful inhibition data points (i.e., solid diamonds) to the lower left of the diagonal lines indicate a synergistic biocide/metabolic inhibitor effect. Combinations of several biocides with nitrite or molybdate resulted in synergistic inhibitory effects. In particular, nitrite plus glutaraldehyde (FIG. 1) or benzalkonium chloride (FIG. 2) and molybdate plus glutaraldehyde (FIG. 6) showed a strong synergistic effect. Nitrite plus Bronopol (FIG. 3) produced a lesser synergistic effect. Nitrite plus cocodiamine (FIG. 4) and molybdate plus benzalkonium chloride (FIG. 7), cocodiamine (FIG. 9), or Bronopol (FIG. 8) produced the smallest synergistic effect. Nitrite plus THPS (FIG. 5) and molybdate plus THPS (FIG. 10) showed a less than additive effect. This less than additive effect with THPS could be an isolated phenomenon for the particular SRB and conditions employed in this study. No combinations tested produced indifferent or antagonistic effects. Thus, all combinations other than those with THPS resulted in better than additive inhibitory effects.

The preferred forms of the invention described above are to be used as illustration only, and should not be used in a limiting sense to interpret the scope of the present invention. Obvious modifications to the exemplary embodiments, set forth above, could be readily made by those skilled in the art without departing from the spirit of the present invention.

The inventors hereby state their intent to rely on the Doctrine of Equivalents to determine and assess the reasonably fair scope of the present invention as it pertains to any apparatus not materially departing from but outside the literal scope of the invention as set forth in the following claims.

What is claimed is:

1. A method of inhibiting sulfide production by sulfate-reducing bacteria (SRB), said method comprising the steps of:

- 5 (a) contacting the SRB with a first concentration of a biocide component, wherein
 said first concentration is less than about 90% of the minimum inhibitory
 concentration (MIC) of the biocide component; and
 (b) contacting the SRB with a second concentration of a metabolic inhibitor
 component, wherein said second concentration is less than about 90% of the
 MIC of the metabolic inhibitor component.

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2. The method of claim 1, wherein at least one of said first and second
concentrations is less than about 50% of its MIC.

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3. The method of claim 3, wherein said first and second concentrations
are both less than about 75% of their respective MICs.

4. The method of claim 1, wherein at least one of said first and second
concentrations is less than about 25% of its MIC.

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5. The method of claim 4, wherein said first and second concentrations
are both less than about 50% of their respective MICs.

6. The method of claim 1, wherein at least one of said first and second
concentrations is less than 20% of its MIC.

25

7. The method of claim 6, wherein said first and second concentrations
are both less than about 35% of their respective MICs.

30

8. The method of claim 1, wherein said second concentration is in the
range of from about 0.1 mM to about 5 mM.

9. The method of claim 8, wherein said first concentration is less than
about 50% of the MIC of the biocide component.

10. The method of claim 1, wherein said biocide component is a combination of more than one individual biocide and/or said metabolic inhibitor component is a combination of more than one individual metabolic inhibitor.

5 11. The method of claim 1, wherein said biocide component comprises substantially no tetrakis hydroxymethyl phosphonium sulfate (THPS).

10 12. The method of claim 1, wherein said biocide component is selected from the group consisting of aldehydes, amine-type compounds, halogenated compounds, sulfur compounds, quaternary phosphonium salts, and combinations of one or more thereof.

15 13. The method of claim 1, wherein said metabolic inhibitor component is selected from the group consisting of nitrite, molybdate, tungstate, selenate, anthraquinone and combinations of one or more thereof.

20 14. The method of claim 1, wherein said biocide component is selected from the group consisting of formaldehyde, glutaraldehyde, acrolein, quaternary amine compounds, cocodiamine, bronopol, 2-2-dibromo-3-nitrilo-propionamide (DBNPA), isothiazolone, carbamates, metronidazole, and combinations of one or more thereof.

25 15. The method of claim 1, wherein said biocide component comprises glutaraldehyde and said metabolic inhibitor component comprises nitrite.

16. The method of claim 1, wherein said biocide component consists essentially of glutaraldehyde and said metabolic inhibitor component consists essentially of nitrite .

30 17. The method of claim 1, wherein step (a) includes directly killing a first portion of the SRB and step (b) includes inhibiting sulfate-reducing growth of a second portion of the SRB without directly killing the second portion of the SRB.

18. The method of claim 1, wherein steps (a) and (b) are performed substantially continuously and said first and second concentrations are average concentrations over time.

5 19. The method of claim 1, wherein steps (a) and (b) are performed intermittently and said first and second concentrations are average concentrations over time.

20. The method of claim 1, wherein steps (a) and (b) are performed simultaneously.

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21. The method of claim 1, further comprising the step of:

(c) prior to steps (a) and (b), combining the biocide component or a precursor of the biocide component and the metabolic inhibitor component or a precursor of the metabolic inhibitor component in a treated medium.

15

22. The method of claim 21, wherein steps (a) and (b) include contacting the SRB with the treated medium.

20 23. The method of claim 21, wherein said treated medium is an aqueous-based medium.

24. The method of claim 21, wherein said treated medium comprises at least about 50% water by weight.

25 25. The method of claim 21, wherein said treated medium comprises in the range of from about 0.1 mM to about 5 mM nitrite.

26. The method of claim 25, wherein said treated medium comprises in the range of from about 0.1 mM to about 5 mM glutaraldehyde.

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27. A method comprising contacting sulfate reducing bacteria (SRB) with a treated medium comprising an aldehyde and a metabolic inhibitor selected from the group consisting of nitrite, molybdate, and combinations thereof, wherein said aldehyde and said metabolic inhibitor are present in the treated medium in an aldehyde to metabolic inhibitor molar ratio in the range of from about 50:1 to about 1:50.

5 28. The method of claim 27, wherein said metabolic inhibitor comprises nitrite.

10 29. The method of claim 28, wherein said biocide comprises glutaraldehyde.

30. The method of claim 27, wherein said aldehyde to metabolic inhibitor molar ratio is in the range of from about 20:1 to about 1:20.

15 31. The method of claim 27, wherein said aldehyde to metabolic inhibitor molar ratio is in the range of from 10:1 to 1:10, wherein said metabolic inhibitor consists essentially of nitrite, and wherein said biocide consists essentially of glutaraldehyde.

20 32. The method of claim 27, wherein said aldehyde and said metabolic inhibitor are each present in the treated medium in a concentration in the range of from about 0.1 mM to about 5 mM.

25 33. The method of claim 32, wherein step (a) is performed substantially continuously and said concentrations of the aldehyde and metabolic inhibitor in the treated medium are average concentrations over time.

30 34. The method of claim 32, wherein step (a) is performed intermittently and said concentrations of the aldehyde and metabolic inhibitor in the treated medium are average concentrations over time.

35. The method of claim 27, wherein said aldehyde and said metabolic inhibitor are each present in the treated medium in a concentration in the range of from about 0.1 mM to about 2 mM.

5 36. The method of claim 27, wherein said aldehyde is present in the treated mixture in a first concentration that is less than about 90% of the minimum inhibitory concentration (MIC) of the aldehyde and said metabolic inhibitor is present in the treated mixture in a second concentration that is less than about 90% of the MIC of the metabolic inhibitor.

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37. The method of claim 36, wherein at least one of said first and second concentrations is less than about 35% of its MIC.

15 38. The method of claim 37, wherein said aldehyde comprises glutaraldehyde and said metabolic inhibitor comprises nitrite.

20 39. The method of claim 27, wherein said aldehyde is present in the treated mixture in a first concentration that is less than about 50% of the minimum inhibitory concentration (MIC) of the aldehyde and said metabolic inhibitor is present in the treated mixture in a second concentration that is less than about 50% of the MIC of the metabolic inhibitor.

40. The method of claim 39, wherein at least one of said first and second concentrations is less than 25% of its MIC.

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41. The method of claim 40, wherein said aldehyde consists essentially of glutaraldehyde and said metabolic inhibitor consists essentially of nitrite.

42. A composition for effectively inhibiting sulfide production by sulfate-reducing bacteria (SRB), said composition comprising:

- (a) a biocide component capable of directly killing a first portion of the SRB, wherein
said biocide component is present in the composition in a first concentration
5 that is less than about 90% of the minimum inhibitory concentration (MIC)
of the biocide component; and
- (b) a metabolic inhibitor component capable of inhibiting the sulfate-reducing growth
of a second portion of the SRB without directly killing the second portion of
the SRB, wherein said metabolic inhibitor component is present in the
10 composition in a first concentration that is less than about 90% of the MIC of
the biocide component.

43. The composition of claim 42, wherein at least one of said first and
second concentrations is less than about 35% of its MIC.

15 44. The composition of claim 43, wherein said first and second
concentrations are both less than about 50% of their respective MICs.

20 45. The composition of claim 42, wherein said at least one of first and
second concentrations is less than 25% of its MIC.

46. The composition of claim 45, wherein said first and second
concentrations are both less than about 35% of their respective MICs.

25 47. The composition of claim 42, wherein said biocide component is a
combination of more than one individual biocide and/or said metabolic inhibitor component
is a combination of more than one individual metabolic inhibitor.

30 48. The composition of claim 42, wherein said biocide component
comprises substantially no tetrakis hydroxymethyl phosphonium sulfate (THPS).

49. The composition of claim 42, wherein said biocide component is
selected from the group consisting of aldehydes, amine-type compounds, halogenated

compounds, sulfur compounds, quaternary phosphonium salts, and combinations of one or more thereof.

5. 50. The composition of claim 42, wherein said metabolic inhibitor component is selected from the group consisting of nitrite, molybdate, tungstate, selenate, anthraquinone, and combinations of one or more thereof.

10. 51. The composition of claim 42, wherein said biocide component is selected from the group consisting of formaldehyde, glutaraldehyde, acrolein, quaternary amine compounds, cocodiamine, bronopol, 2-2-dibromo-3-nitrilo-propionamide (DBNPA), isothiazolone, carbamates, metronidazole, and combinations of one or more thereof.

15. 52. The composition of claim 42, wherein said biocide component comprises glutaraldehyde and said metabolic inhibitor component comprises nitrite.

53. The composition of claim 42, wherein said biocide component consists essentially of glutaraldehyde and said metabolic inhibitor component consists essentially of nitrite.

20. 54. The composition of claim 42, further comprising at least about 2% by weight water.

55. The composition of claim 42, further comprising at least 50% by weight water.

25

56. A composition comprising:

- (a) an aldehyde; and
- (b) a metabolic inhibitor selected from the group consisting of nitrite, molybdate, and combinations thereof,

5 wherein said aldehyde and said metabolic inhibitor are present in the composition in an aldehyde to metabolic inhibitor molar ratio in the range of from about 50:1 to about 1:50.

57. The composition of claim 56, wherein said metabolic inhibitor
10 comprises nitrite.

58. The composition of claim 57, wherein said biocide comprises glutaraldehyde.

15 59. The composition of claim 56, wherein said aldehyde to metabolic inhibitor molar ratio is in the range of from about 20:1 to about 1:20.

20 60. The composition of claim 56, wherein said aldehyde to metabolic inhibitor molar ratio is in the range of from 10:1 to 1:10, wherein said metabolic inhibitor consists essentially of nitrite, and wherein said biocide consists essentially of glutaraldehyde.

61. The composition of claim 56, wherein said aldehyde and said metabolic inhibitor are each present in the composition in a concentration in the range of from about 0.1 mM to about 5 mM.

25 62. The composition of claim 56, wherein said aldehyde and said metabolic inhibitor are each present in the composition in a concentration in the range of from about 0.1 mM to about 2 mM.

30 63. The composition of claim 56, wherein said aldehyde comprises glutaraldehyde and said metabolic inhibitor comprises nitrite.

64. The composition of claim 56, wherein said aldehyde consists essentially of glutaraldehyde and said metabolic inhibitor consists essentially of nitrite.

ABSTRACT

Biogenic sulfide production is synergistically inhibited by treating sulfate-reducing bacteria (SRB) with a biocide and a metabolic inhibitor. The biocide 5 directly kills a first portion of the SRB. The metabolic inhibitor inhibits sulfate-reducing growth of a second portion of the SRB without directly killing the second portion of the SRB. The treatment of SRB with both a biocide and a metabolic inhibitor provides effective biogenic sulfide inhibition at significantly lower concentrations than would be required if the biocide or metabolic inhibitor was used alone.

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DECLARATION, POWER OF ATTORNEY, AND PETITION

To The Commissioner of Patents and Trademarks:

I, Gary E. Jenneman, declare:

that my residence, mailing address and citizenship are as stated below;

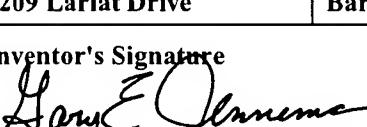
that I have reviewed and understand the contents of the attached specification and claims and I verily believe I, Anne Greene, and Gerrit Voordouw are the original, first, and joint inventors or discoverers of the invention or discovery in **INHIBITION OF BIOGENIC SULFIDE PRODUCTION VIA BIOCIDE AND METABOLIC INHIBITOR COMBINATION** described and claimed therein and for which a patent is sought;

I acknowledge my duty to disclose to the U.S. Patent and Trademark Office all information known to me to be material to patentability as defined in Title 37 Code of Federal Regulations § 1.56;

and I hereby appoint Jeffrey R. Anderson, Registration No. 42,263; Anne E. Brookes, Registration No. 33,066; Ryan N. Cross, Registration No. 33,861; Beverly M. Dollar, Registration No. 32,870; Gary L. Haag, Registration No. 35,529; Lynda S. Jolly, Registration No. 32,736; Kim S. Manson, Registration No. 42,839; Patricia A. Meier, Registration No. 43,798; Bronwyn A. Welvaert, Registration No. 52, 350; and David W. Westphal, Registration No. 32,673; who comprise the firm of **Richmond, Hitchcock, Fish & Dollar, P. O. Box 2443, Bartlesville, Oklahoma, 74005**, Registration No. 20,286; and Kameron D. Kelly, Registration No. 44,181; my representatives with full power of substitution and revocation, to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith.

I declare further that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements are made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of this application or any patent issuing thereon.

Wherefore I pray that Letters Patent be granted to ConocoPhillips Company, a corporation organized and existing under the laws of the State of Delaware and domiciled and having an office at Bartlesville, Oklahoma, as assignee, for the invention or discovery described and claimed in the foregoing specification and claims, and I hereby subscribe my name to the foregoing specification and claims, declaration, power of attorney and this petition.

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DECLARATION, POWER OF ATTORNEY, AND PETITION

To The Commissioner of Patents and Trademarks:

I, Anne Greene, declare:

that my residence, mailing address and citizenship are as stated below;

that I have reviewed and understand the contents of the attached specification and claims and I verily believe I, Gary E. Jenneman, and Gerrit Voordouw are the original, first, and sole inventors or discoverers of the invention or discovery in **INHIBITION OF BIOGENIC SULFIDE PRODUCTION VIA BIOCIDE AND METABOLIC INHIBITOR COMBINATION** described and claimed therein and for which a patent is sought;

I acknowledge my duty to disclose to the U.S. Patent and Trademark Office all information known to me to be material to patentability as defined in Title 37 Code of Federal Regulations § 1.56;

and I hereby appoint Jeffrey R. Anderson, Registration No. 42,263; Anne E. Brookes, Registration No. 33,066; Ryan N. Cross, Registration No. 33,861; Beverly M. Dollar, Registration No. 32,870; Gary L. Haag, Registration No. 35,529; Lynda S. Jolly, Registration No. 32,736; Kim S. Manson, Registration No. 42,839; Patricia A. Meier, Registration No. 43,798; Bronwyn A. Welvaert, Registration No. 52, 350; and David W. Westphal, Registration No. 32,673; who comprise the firm of **RICHMOND, HITCHCOCK, FISH & DOLLAR, P. O. Box 2443, Bartlesville, Oklahoma, 74005**, Registration No. 20,286; and Kameron D. Kelly, Registration No. 44,181; my representatives with full power of substitution and revocation, to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith.

I declare further that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements are made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of this application or any patent issuing thereon.

Wherefore I pray that Letters Patent be granted for the invention or discovery described and claimed in the foregoing specification and claims, and I hereby subscribe my name to the foregoing specification and claims, declaration, power of attorney and this petition.

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DECLARATION, POWER OF ATTORNEY, AND PETITION

To The Commissioner of Patents and Trademarks:

I, Gerrit Voordouw, declare:

that my residence, mailing address and citizenship are as stated below;

that I have reviewed and understand the contents of the attached specification and claims and I verily believe I, Gary E. Jenneman, and Anne Greene are the original, first, and joint inventors or discoverers of the invention or discovery in **INHIBITION OF BIOGENIC SULFIDE PRODUCTION VIA BIOCIDE AND METABOLIC INHIBITOR COMBINATION** described and claimed therein and for which a patent is sought;

I acknowledge my duty to disclose to the U.S. Patent and Trademark Office all information known to me to be material to patentability as defined in Title 37 Code of Federal Regulations § 1.56;

and I hereby appoint Jeffrey R. Anderson, Registration No. 42,263; Anne E. Brookes, Registration No. 33,066; Ryan N. Cross, Registration No. 33,861; Beverly M. Dollar, Registration No. 32,870; Gary L. Haag, Registration No. 35,529; Lynda S. Jolly, Registration No. 32,736; Kim S. Manson, Registration No. 42,839; Patricia A. Meier, Registration No. 43,798; Bronwyn A. Welvaert, Registration No. 52, 350; and David W. Westphal, Registration No. 32,673; who comprise the firm of **Richmond, Hitchcock, Fish & Dollar, P. O. Box 2443, Bartlesville, Oklahoma, 74005**, Registration No. 20,286; and Kameron D. Kelly, Registration No. 44,181; my representatives with full power of substitution and revocation, to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith.

I declare further that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements are made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of this application or any patent issuing thereon.

Wherefore I pray that Letters Patent be granted for the invention or discovery described and claimed in the foregoing specification and claims, and I hereby subscribe my name to the foregoing specification and claims, declaration, power of attorney and this petition.

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